

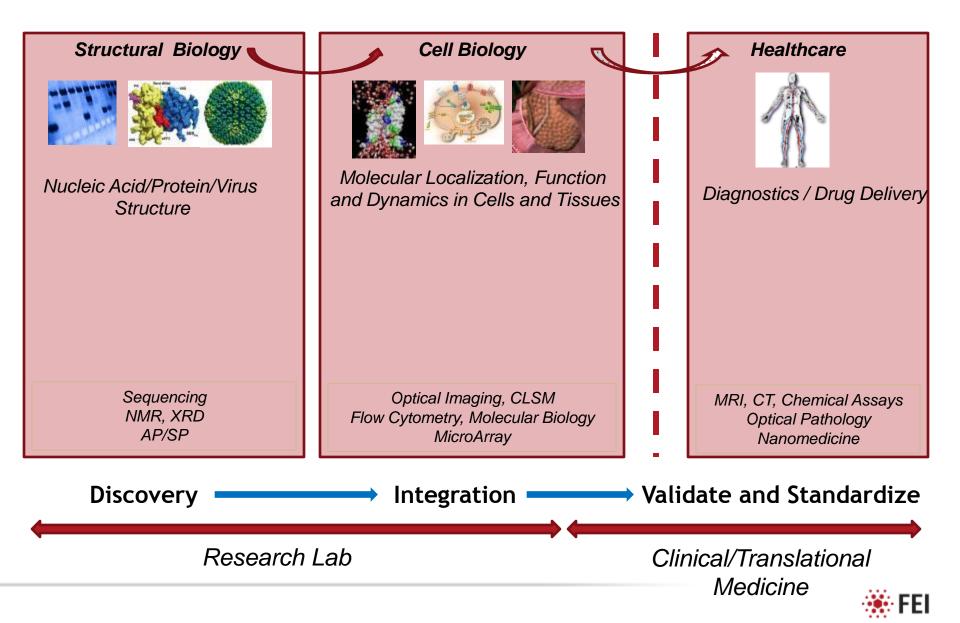


Correlative Microscopy: Bridging the Gap Between Light Microscopy and Electron Microscopy

Wim Voorhout

Advanced Technologies for Life Sciences Institut Pasteur 14-15 September, 2010

Life Sciences Landscape



Imaging landscape

Resolution -

Organs Cells in Tissue Molecules in Cells In vivo imaging Light microscopy Electron microscopy Resolution + System Biology Cellomics + Genomics/Proteomics

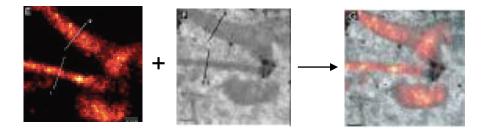


Correlative Microscopy - Motivation

LM and EM are complementary techniques:

- LM for identifying locations of interest and dynamic events using fluorescent tags
- TEM for zooming in to nm resolution to provide cellular context

As recently reported in Nature Methods, Jennifer Lippincott-Schwartz states: "Unorthodox super-resolution microscopy discoveries will also need support from electron microscopy. The latter is especially important as it provides the needed nanometer-scale resolution of cell ultra structure to correlate with super resolution images."





Correlative Microscopy

From macromolecular structure to it's cellular context and v.v.

Available technologies

Light Microscopy

Scanning Electron Microscopy

Transmission Electron Microscopy

Scanning Probe Microscopy

XRD

NMR



Correlative SEM - Fluorescent microscopy

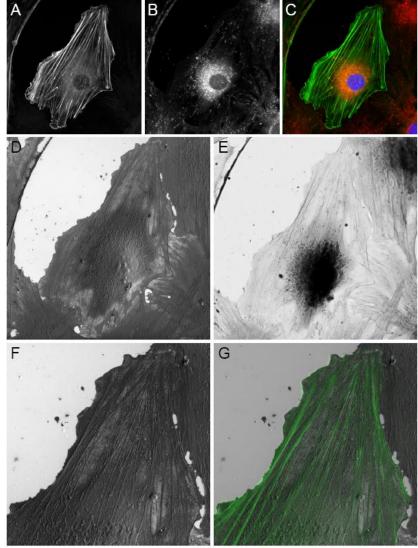
Mouse embryonic fibroblasts expressing YFP-B-actin, grown on Indium-Tin Oxide slides

Correlate

- Fluorescent Light Microscopy
- Secondary Electron
- Backscatter Electron

H. Pluk, J. Fransen University Nijmegen, Netherlands

Pluk et al. <u>J Microsc</u>. 2009; 233(3): 353-63

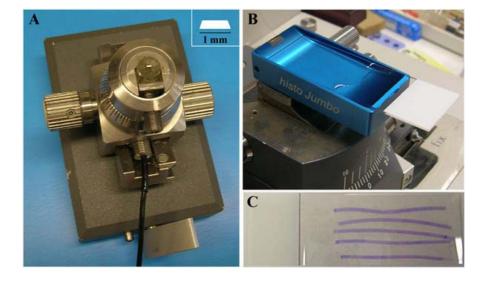




Array Tomography

Array Tomography: A New Tool for Imaging the Molecular Architecture and Ultrastructure of Neural Circuits

¹ Department of Molecular and Cellular Physiology, Stanford University, Stanford, CA 94305, USA *Correspondence: kmicheva@stanford.edu (K.D.M.), sjsmith@stanford.edu (S.J S.) DOI 10.1016/j.neuron.2007.06.014



Reprinted from: Neuron 55, 25 – 36, 2007

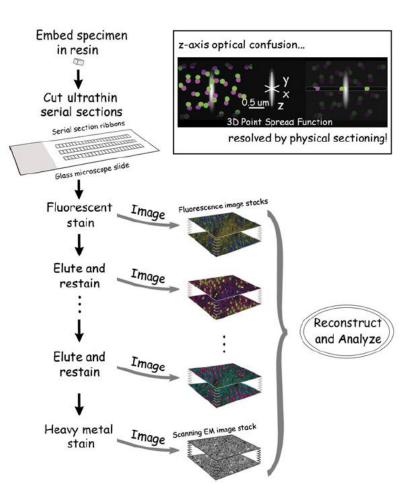
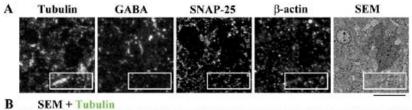


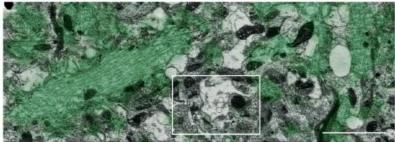
Figure 1. Schematic Representation of the Array Tomography Method



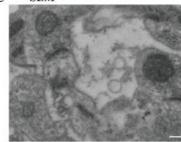
Kristina D. Micheva^{1,*} and Stephen J Smith^{1,*}

Array Tomography





C SEM



D Tubulin + GABA

β-actin + SNAP-25

S

S

G

sp

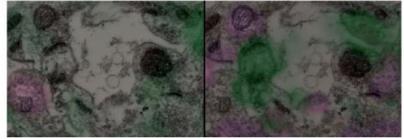


Figure 7. Demonstration of Naturally Excellent Registration of Light and Electron Microscopic Imaging of an Individual Specimen by Array Tomography

(A) The same region of a 70 nm section from the mouse cerebral cortex is shown as immunostained for tubulin, GABA, SNAP-25, and β -actin and imaged in the SEM. Scale bar, 10 μ m.

(B) The boxed region in (A) is imaged at a higher magnification in the SEM, and the corresponding immunofluorescent labeling for tubulin (green) is overlayed. Scale bar, $2 \,\mu$ m.

(C) A higher-magnification SEM image of the boxed region in (B) and a schematic map of the same region: G-glia, S-presynaptic bouton, sp-spine. Scale bar, 0.5 μ m.

(D) Immunofluorescence for tubulin and GABA, and β-actin and SNAP-25 superimposed on the SEM image in (C). Scale bar, 0.5 μm.

Reprinted from: Neuron 55, 25 – 36, 2007



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Transmission Electron Microscopy

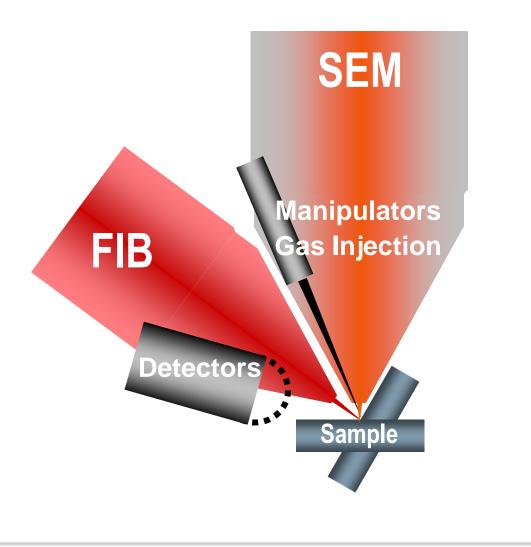
Scanning Probe Microscopy

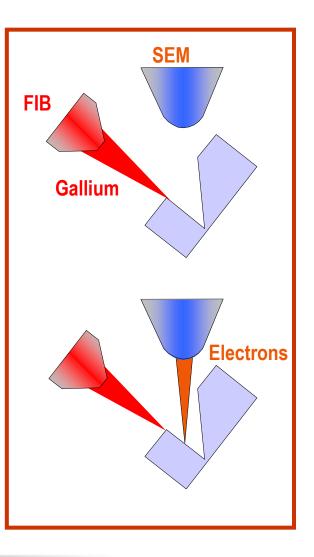
XRD

NMR



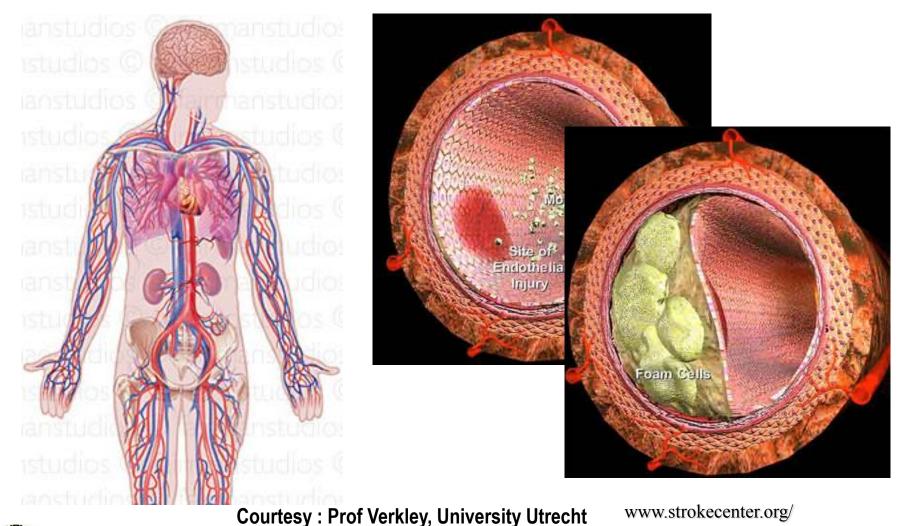
What is a DualBeam?







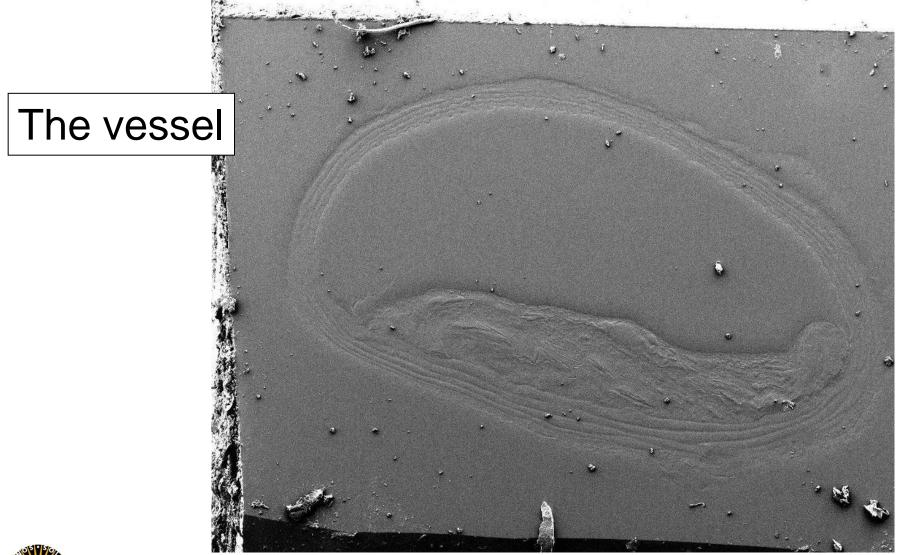
Atherosclerosis : Needle in the haystack problem







Atherosclerosis : Needle in the haystack problem

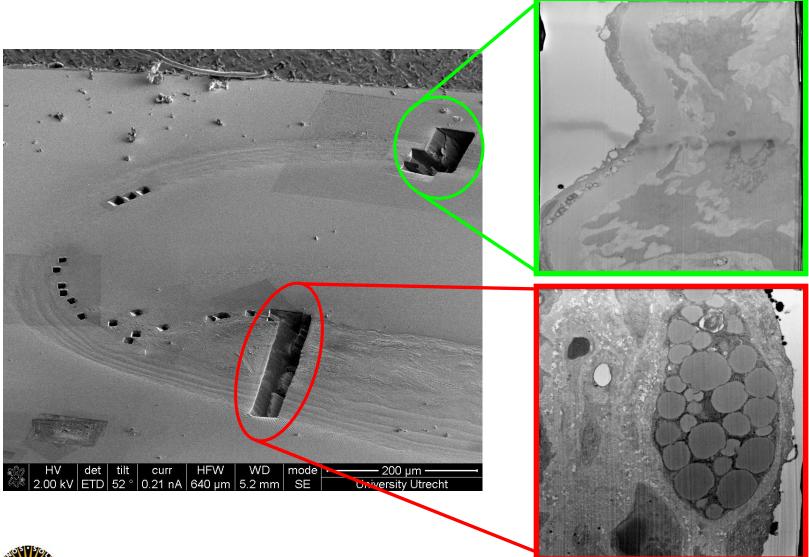




Courtesy : Prof Verkley, University Utrecht



Atherosclerosis : Needle in the haystack problem

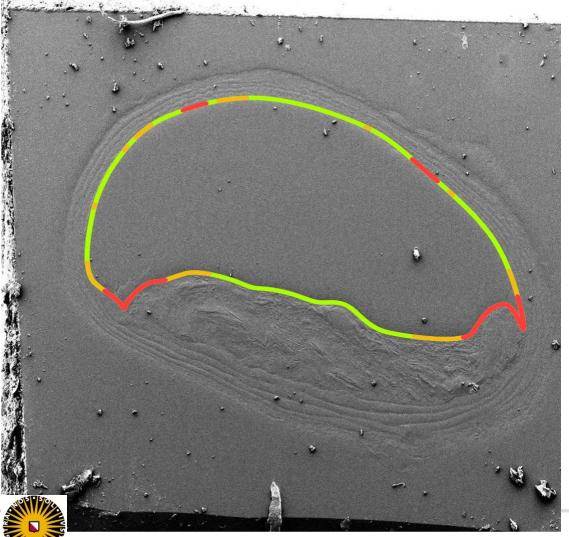




Courtesy : Prof Verkley, University Utrecht



Spatial Density of ICAM specific label at inner blood vessel



high number of gold particles could be indicative for early stage plaque development low number indicates healthy tissue

Journal of Microscopy, Vol. 235, Pt 3 2009, pp. 336-347



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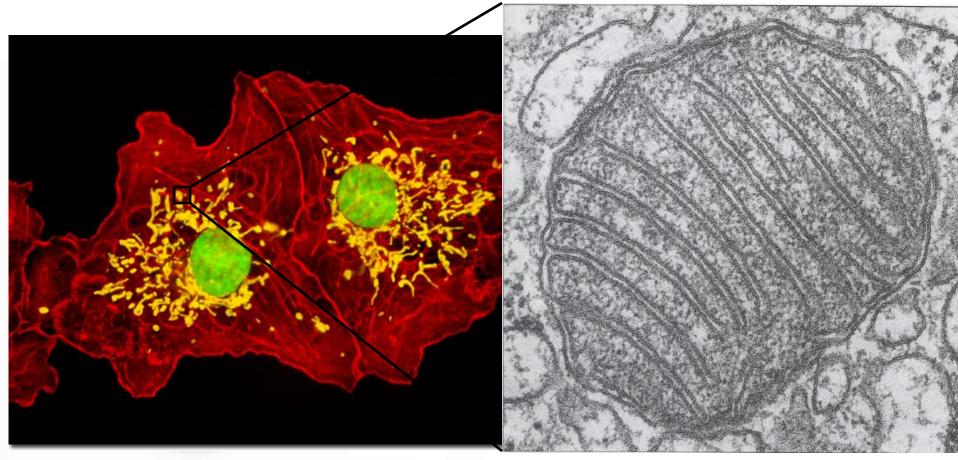
Scanning Probe Microscopy

XRD

NMR



Correlative Microscopy - Leveraging the Strengths of Both LM and EM



Olympus Inverted Light Microscope

FEI Tecnai Spirit TEM



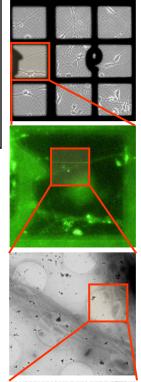
Correlative Microscopy

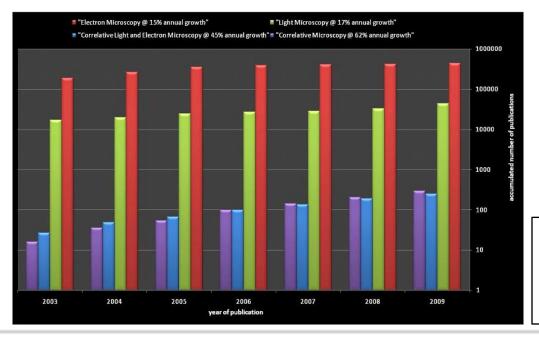
Light Microscopy (widely used in biology)

- Large field of view / overview
- Live cell imaging
- Limited or no sample preparation
- Combination with fluorescent labels (GFP..)
- Resolution ~ 200 nm

Electron Microscopy

- Excellent spatial resolution with small field of view
- Provides the cellular context
- Extensive specimen preparation, especially for labeling techniques

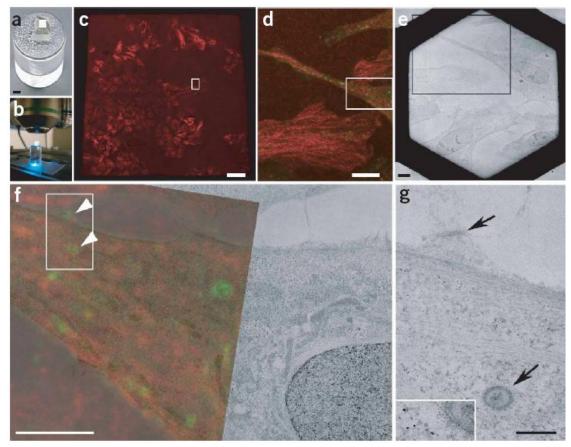




Growth rates for correlative papers are highest. EM=15%, LM=17%, CLEM=45%, CM=62%. A clear trend towards correlative microscopy.



Correlative Microscopy (manual)



Confocal block-face imaging

Ultrathin sectioning

TEM

Correlated light and electron microscopic imaging of multiple endogenous proteins using Quantum dots

Ben N G Giepmans, Thomas J Deerinck, Benjamin L Smarr, Ying Z Jones & Mark H Ellisman

NATURE METHODS | VOL.2 NO.10 | OCTOBER 2005 | 743

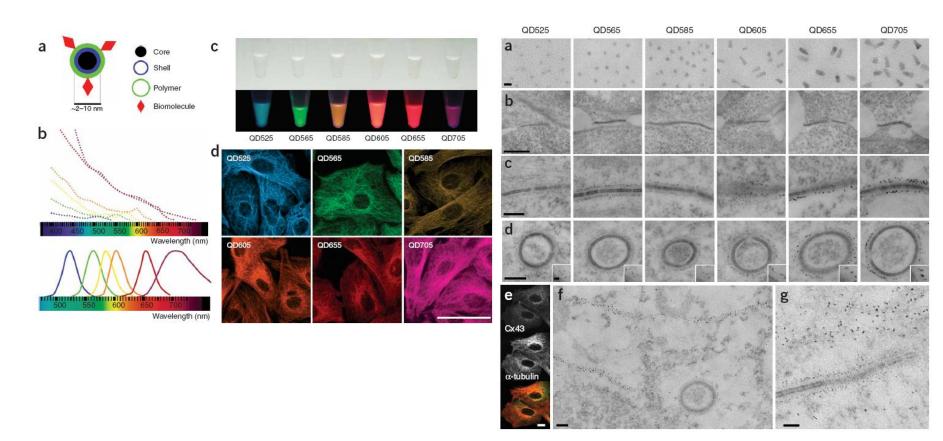


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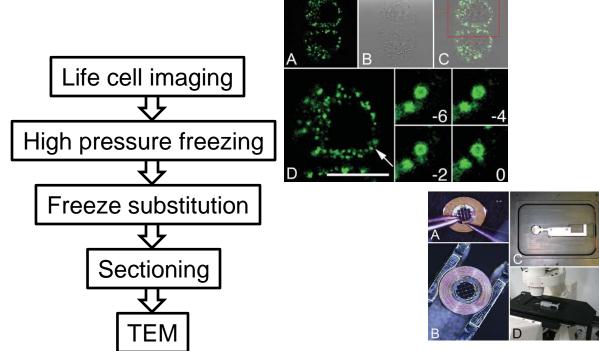
NATURE METHODS | VOL.2 NO.10 | OCTOBER 2005 | 743





Correlative workflow

Example Bristol (Paul Verkade)

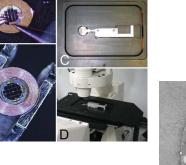


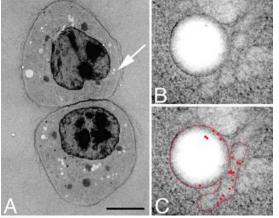


P. VERKADE

Moving EM: the Rapid Transfer System as a new tool for correlative light and electron microscopy and high throughput for high-pressure freezing

Journal of Microscopy, Vol. 230, Pt 2 2008, pp. 317–328

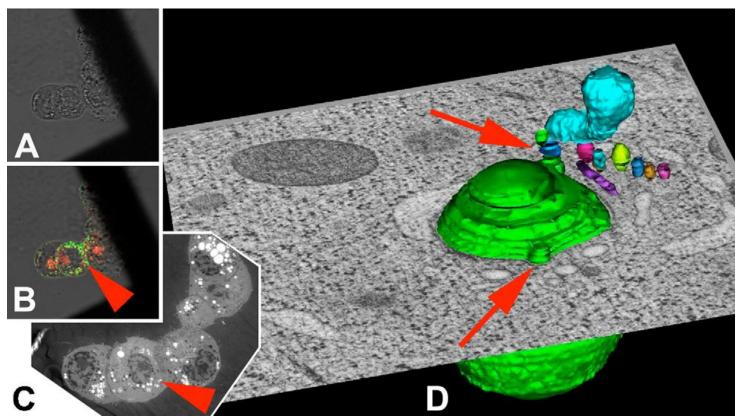






Correlative workflow

• Example Bristol (Paul Verkade)

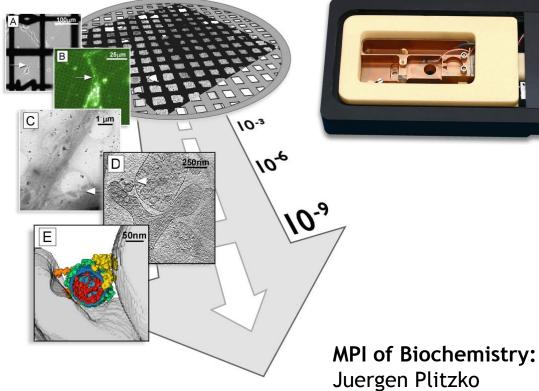


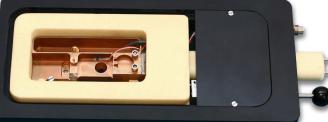
E. Brown et al. / Seminars in Cell & Developmental Biology 20 (2009) 910-919





Cryo-correlative workflow













Network of Excellence (NoE) in 3DEM

Alexander Rigort

Anna Sartori (Pasteur)

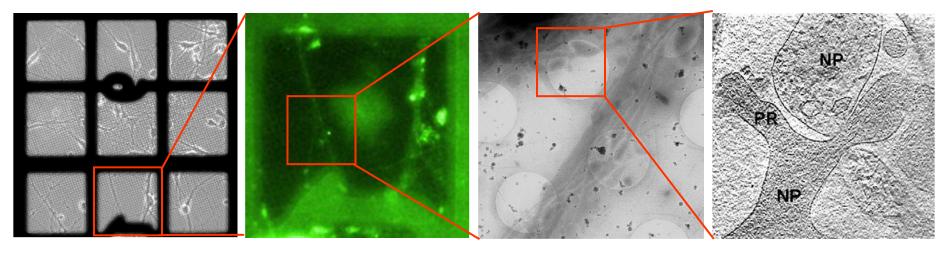
Andrew Leis

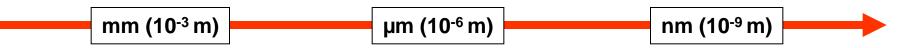


Cryo-correlative Microscopy

Correlative microscopy - characteristics

- Examining one and the same sample by both light and electron microscopy
- LM provides a survey over large cellular landscapes
- FM allows positive identification of features of interest
- EM and ET permits zooming in on such features at much higher resolution



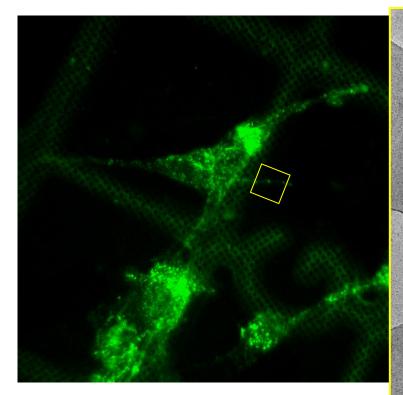


Example given: Cultured neurons grown on EM gold finder grids

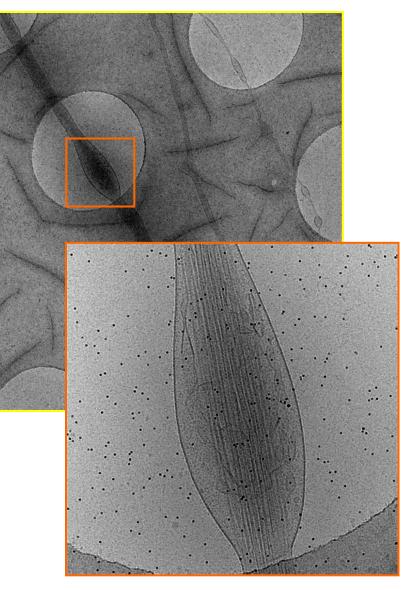




Correlating FM and cryo-ET: Cryo-FM features

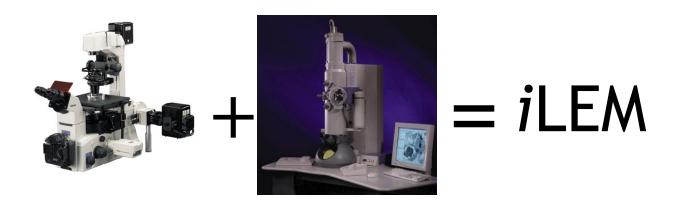


HFF (Human Foreskin Fibroblasts) live immunolabelling with Ab conjugated to Alexa488





Integrated Correlative Microscopy



Hans C. Gerritsen, Sasha Agronskaia, Abraham J. Koster, Arie J. Verkleij



Universiteit Utrecht



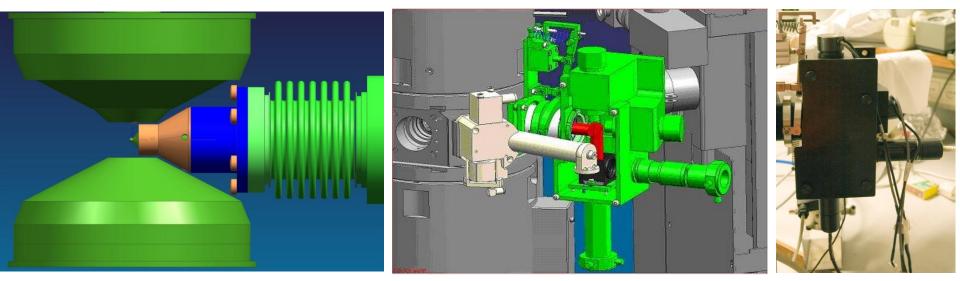
FEI contributed to the design and construction of a prototype





Correlative Microscopy

Integrated Correlative Light Electron Microscopy (ILEM)



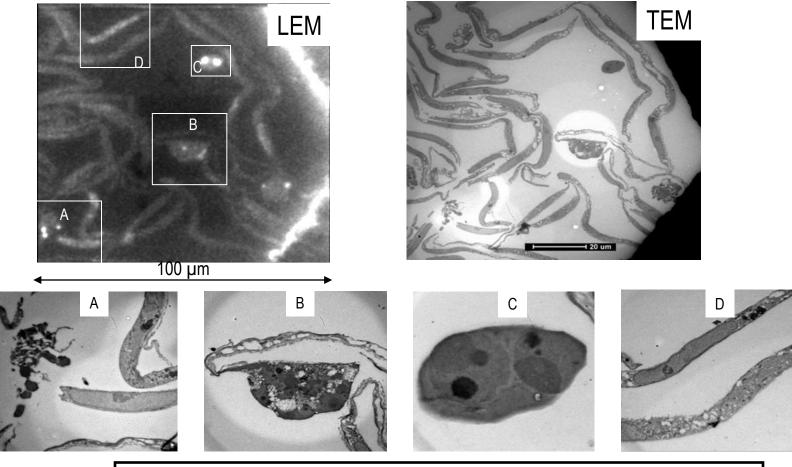
- Optimal Navigation Tool for correlative microscopy
 - Faster correlation between LM and EM
 - Less potential contamination (cryo)
 - Less photo-bleaching of fluorescent labels (vacuum)
 - No compromise on EM performance, slight decline in LM resolution
- Prototype ILEM up and running on Tecnai 12 at Utrecht University
 - Paper published in J. Structural Biology (Volume 164, 2008, 183-189)
- Second prototype was recently installed at Leiden University
- First application results have been obtained (next slides)
- Funded by STW





Correlative Microscopy - ILEM

UVC stressed HUVEC cells, staining: γ -H2AX with Alexa488 & gold



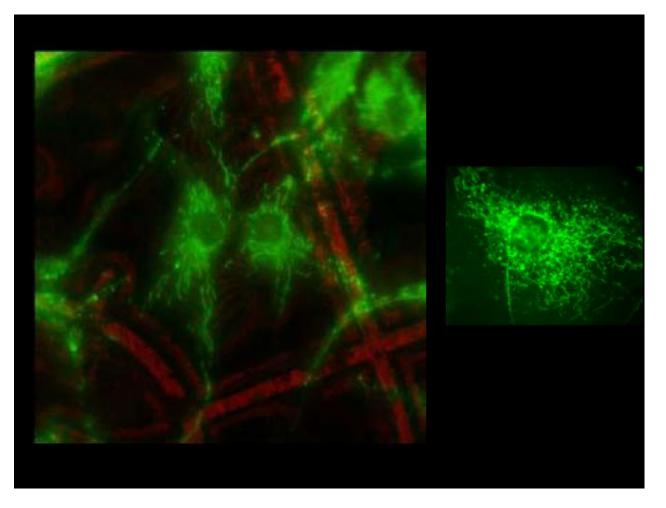


FEM

M.A. Karreman et al., Biology of the Cell, (2009) 101, 287-299



Cryo-correlative microscopy

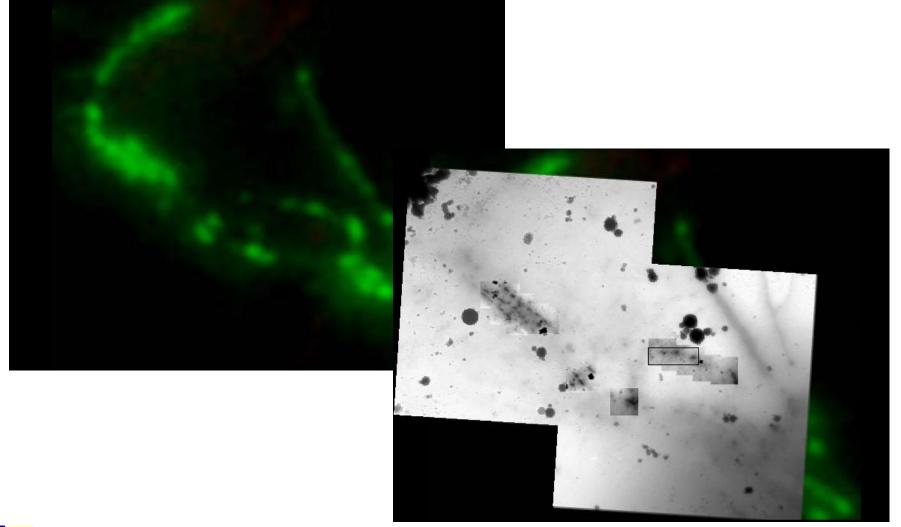


Courtesy: Linda van Driel and Bram Koster, Leiden University, The Netherlands





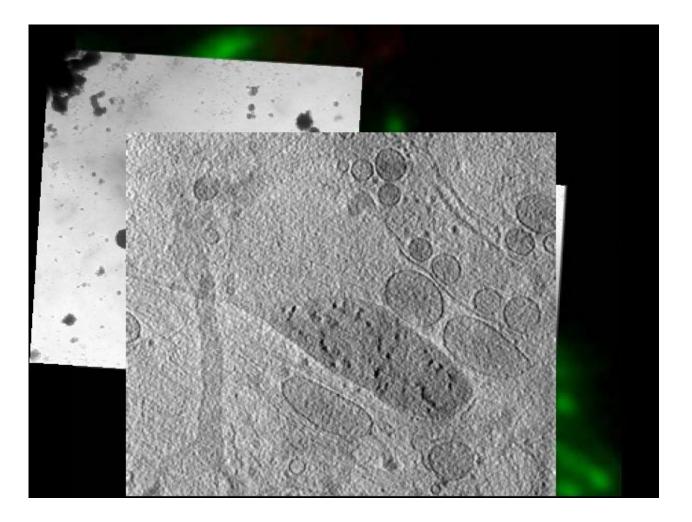
Cryo-correlative microscopy







Cryo-correlative microscopy





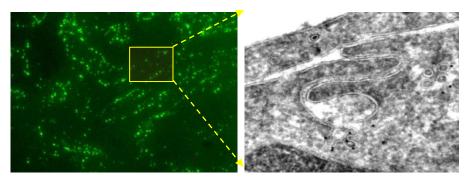


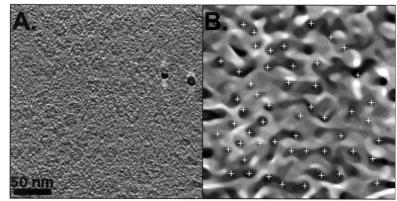
Biology Challenges

- Link MRI/CT Medical Data to ultrastructural data
- Link Light Microscopy data to EM Data
- Link Cellular Architecture context to localization of specific **Macromolecules**
- Link X-ray Structures to EM Data
- With minimal Artifacts (Close to living state)
- In a significant volume

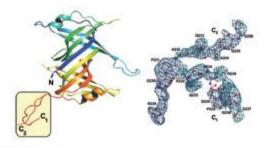
The key is correlation!

Biomarker Localization: LM Biomarker Validation: EM





Böhm J et al. PNAS 2000;97:14245-14250





Acknowledgements

Molecular Biophysics UU

Hans Gerritsen Sasha Agronskia

Cell Biology UU

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Molecular Cell Biology LUMC

Bram Koster Jack Valentijn Linda van Driel

MPI of Biochemistry

Juergen Plitzko Alexander Rigort Tim Laugks Andrew Leis Anna Sartori (Pasteur)

Dept. Of Biochemistry, Bristol

Paul Verkade Edward Brown



Max Planck Institute of Biochemistry



6th EU Framework



